

Analysing the Impact of Broiler Line and Egg Preservation Durations on Immunoglobulin Levels and Some Egg Quality Indicators

Ali N. Zaki¹, Hashim Hadi Al-Jebory², Mohammed Khalil Ibrahim Al-Saedi³

¹Department of Animal production - College of Agriculture/University of sumer, Dhi Qar, Iraq

²Department of animal production, agriculture college- Al-Qasim green university, Iraq

³Department of Environmental - College of Environmental Sciences Al-Qasim Green University, Iraq

ABSTRACT: The basis of the poultry industry is the presence of parent flocks, and the lack of this causes the eggs to be transported for long periods and long distances, reducing the quality of hatching eggs and producing poor-quality, low-weight chicks. Therefore, the current study investigates the effect of different storage periods for different types of broiler chickens on the internal immunity of the egg and some qualitative characteristics of stored eggs. Three lines of broiler were used from three hatcheries (Ross 308-1: Ross 308-2: Cobb 508); these lines of eggs were imported from Türkiye and the Netherlands to Iraq, and all eggs were storage for four periods (15, 20, 25 and, 30 days) at seven c°. The results showed a significant ($P \leq 0.05$) increase for (line- 2) in total IgY at 25 days, and line three increased significantly ($P \leq 0.05$) in the HU unit at 25 days too. Meanwhile, there is no significant difference between periods in egg weight loss (Table 2), albumin pH, relative albumin weight, and yolk weight. It is concluded from the study's results that any storage duration may affect the egg's internal immunity and its qualitative characteristics. Thus, it may negatively affect the hatching rate and characteristics of the hatched chicks.

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Corresponding Author:
Hashim Hadi Al-Jebory

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INTRODUCTION

Storing hatching eggs, regardless of the level of storage temperature, leads to the deterioration of the quality traits of the eggs and, thus, the deterioration of the hatching rate and the quality of the hatched chicks. This results from the lack of availability of broiler breeder stock in Iraq, which forces the owners of fields and hatcheries to import eggs from outside Iraq, which causes them to be stored for long periods, which made many studies address early nutrition to improve the level of nutrients within the eggs and increase their availability to the embryo, which improves the weight and quality of the hatched chicks (Kadhim et al., 2021; Zaki and Al-jebory, 2021; Al-Saedi et al., 2022; AL-ASEEDI et al., 2023; Al-Jaryan et al., 2023; AL-JEBORY et al., 2024). Therefore, using the proper technique while storing eggs is crucial to maintaining their quality; eggs are degraded mainly by temperature, humidity, air movement, handling, and length of storage (Bell, 1996; Samli et al., 2005). Because quality deterioration happens more quickly at hot temperatures than cold temperatures during storage, regulating storage temperature can significantly delay the decline of interior quality in eggs (Zeidler, 2002). The pH and quality of the albumen are the primary differences between fresh and preserved eggs (Walsh et al., 1995). The egg albumen's pH rises during storage, which is connected to the albumen's declining quality or Haugh unit (Jones et al., 2002). An AA egg has a robust and thick albumen, a tiny air cell, and no meat or blood stains in the yolk or albumen. The fresh albumen has the weakest buffering capacity between pH 7.0 and 9.0. Additional parameters influencing the rise in albumen pH are temperature, length of storage, gaseous conditions in the storage room, and eggshell conductivity (Akter et al., 2014). Inadequate levels of IgY in the egg yolk are likely to heighten the susceptibility of the hatching chick to infections. Conversely, extended periods of egg storage will lead to a reduction in immunoglobulin content in the yolk as a consequence of chemical alterations in the egg. IgY is transmitted from the hen to the egg to safeguard the offspring from infections until the chick's immune system has reached a level of maturity that permits the young animal to produce adequate quantities of antibodies (Carlander et al., 2003). Thus, the objective of the present work was to investigate the alterations that take place in hatching eggs throughout various storage durations and for different lines of broiler breeds.

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MATERIALS AND METHODS

In this study, eggs were collected from three sites (hatches): Ross 308-1 (first local), Ross 308-2 (second local), and Cobb 508 (third local). The laboratory work was conducted in the College of Agriculture/Al-Qasim Green University's laboratory from 1 February until the end of April. Studied traits were the total IgY according to (Carlander et al., 2003). Egg weight loss and albumin pH according to (Akter et al., 2014). The HU unites and relative weight of albumin and yolk, according to (Aljebory and Naji, 2021). The SAS (2012) was used for data analysis, and the Duncan multiple ranges test (Duncan, 1955) by model $Y_{ij} = \mu + \tau_i + \epsilon_{ij}$.

RESULTS AND DISCUSSION

The effect of study in yolk total IgY shown in (table 1), noted a not significant difference between periods except for at (25 days) significant ($P \leq 0.05$) increase for (line- 2) compared to (line- 3), in the same period (table 3) the line three increase significantly ($P \leq 0.05$) compared to line one and two in HU unit. There was no significant difference between periods in egg weight loss (Table 2), albumin pH (Table 4), and relative albumin weight (table 5) and, relative yolk weight (table 6).

From the hen to the egg, IgY is actively transported (Rose & Orlans, 1981), according to Bollen and Hau (1997), there is a continuous transfer of IgY from serum to yolk during oocyte development, as evidenced by the consistent IgY concentration in the growing yolk, there is also a movement in feed to use fewer antibiotics, which could raise the risk of illness, this study found a continuous decrease in IgY over time, since the yolk is the chicken's primary source of antibodies, it is likely that chicks hatched from eggs with low IgY concentrations will have less protection, moreover, vaccination of the hens is likely to be unsuccessful and result in poor protection for the chick, these animals have the potential to infect the rest of the flock after contracting the virus, our findings indicate that there are differences in IgY concentrations between individual eggs in the first and second lines as well as between lines, the decrease in the level of immune antibodies may be due to chemical changes that occur over the period of storage and due to the transfer of albumin to yolk and from yolk to albumin, which causes a deterioration in the egg's immunity (Scottand Silversides, 2000: Samli et al., 2005: Akyurek and Okur, 2009). Variations in temperature can influence the pH of egg albumen, leading to increased levels of evaporation from the eggs. Further studies by Moula et al. (2009) and Silversides and Budgell (2004) have also documented this rise in albumen pH during storage. In contrast, our investigation did not observe a statistically significant impact on pH over various storage durations (Table 4). Furthermore, the storage of eggs resulted in elevated pH levels compared to fresh eggs. This phenomenon can be attributed to the process of evaporation and diffusion of carbon dioxide from eggs. The Haugh unit of eggs declined after 25 days of storage, indicating that storage temperatures might exert an influence on egg HU. Increased storage temperatures promote the breakdown of the ovomucin-lysozyme complex, resulting in a reduction in the HU of the stored eggs (Morais et al., 1997).

Table 1: Effect of line broiler on total IgY in different storage periods

Storage Periods				
Line	Fifteen days	Twenty days	Twenty five days	Thirty days
Ross- 308- 1	2.41± 0.28	1.95± 0.06	1.45 ± 0.03 ab	1.16± 0.03
Ross 308- 2	2.47± 0.38	1.79± 0.21	1.57± 0.12 a	1.08± 0.08
Cobb 508- 3	1.62± 0.01	1.56± 0.03	1.41± 0.02 b	1.14± 0.15
Significant	NS	NS	*	NS

NS: Not significant.
Means with different litter significantly at level * ($P \leq 0.05$).

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Table 2: Effect of line broiler on egg weight loss in different storage periods

Storage Periods				
Line	Fifteen days	Twenty days	Twenty five days	Thirty days
Ross- 308- 1	1.46± 0.03	2.47± 0.06	3.11± 0.01	3.83± 0.20
Ross 308- 2	1.46± 0.08	2.50± 0.13	3.17± 0.03	3.73± 0.04
Cobb 508- 3	1.32± 0.09	2.46± 0.01	3.01± 0.01	3.76± 0.04
Significant	NS	NS	NS	NS
NS: Not significant.				

Table 3: Effect of line broiler on HU unite in different storage periods

Storage Periods				
Line	Fifteen days	Twenty days	Twenty five days	Thirty days
Ross- 308- 1	79.61± 0.49	73.85± 0.27	70.69± 0.39 b	65.77± 0.53
Ross 308- 2	79.62± 0.93	74.59± 1.53	70.08± 0.17 b	65.45± 2.13
Cobb 508- 3	80.12± 0.02	76.59± 1.37	73.12± 0.57 a	68.02± 0.13
Significant	NS	NS	*	NS
NS: Not significant. Means with different litter significantly at level * (P≤0.05).				

Table 4: Effect of line broiler on albumin pH in different storage periods

Storage Periods				
Line	Fifteen days	Twenty days	Twenty five days	Thirty days
Ross- 308- 1	8.48± 0.17	8.89± 0.02	8.87± 0.02	8.85± 0.11
Ross 308- 2	8.41± 0.27	8.82± 0.06	8.73± 0.16	8.80± 0.12
Cobb 508- 3	8.51± 0.05	8.75± 0.19	8.73± 0.02	8.89± 0.01
Significant	NS	NS	NS	NS
NS: Not significant.				

Table 5: Effect of line broiler on relative albumin weight in different storage periods

Storage Periods				
Line	Fifteen days	Twenty days	Twenty five days	Thirty days
Ross- 308- 1	56.72± 1.39	55.04± 1.37	53.24± 1.88	52.65± 1.90
Ross 308- 2	58.23± 1.98	57.20± 1.42	55.56± 1.31	54.06± 1.08
Cobb 508- 3	56.94± 2.16	54.32± 2.15	53.34± 1.78	52.63± 1.79
Significant	NS	NS	NS	NS
NS: Not significant.				

Table 6: Effect of line broiler on relative yolk weight in different storage periods

Storage Periods				
Line	Fifteen days	Twenty days	Twenty five days	Thirty days
Ross- 308- 1	21.16± 0.99	22.78± 0.91	23.14± 1.09	23.71± 0.92
Ross 308- 2	20.77± 0.59	21.56± 0.43	22.46± 0.49	22.83± 0.27
Cobb 508- 3	21.57± 0.59	22.09± 0.22	22.99± 0.90	24.01± 0.85
Significant	NS	NS	NS	NS
NS: Not significant.				

CONCLUSION

Previous literature has shown a significant change in all quality egg traits. However, under conditions of high temperature, we also have an arithmetic change in the studied characteristics and a significant decrease in antibodies and Hu unit. All of this negatively affects the hatching rate, the weight of the hatched chicks, and their qualitative characteristics, and this effect continues when the chicks are transported. The breeding halls and the breeder are harmed because of the poor vitality of the chicks; it also results in the chicks not responding to the vaccine programs due to the deterioration of their immune status. Therefore, it is recommended that hatching eggs should be stored for at most five days at a maximum.

REFERENCES

1. Akter, T., A. Kasim, H. Omar, and A.Q. Sazili. 2014. Effect of storage time and temperature on the quality characteristics of chicken eggs. *Journal of Food, Agriculture & Environment*.12 (3&4): 87-92.
2. Akyurek, H. and Okur, A. A. 2009. Effect of storage time, temperature and hen age on egg quality in free-range layer hens. *Journal of Animal and Veterinary Advances* 8:1953-1958.

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3. Al-Jaryan., I.L., H.H. Al-Jebory and M.K.I. Al-Saeed. 2023. Effect of early feeding with different levels of anthocyanins in hatching, phenotypical and physical traits of hatching broiler chicks (Ross 308). *Research Journal of Agriculture and Biological Sciences*, 15(1): 7-13.DOI: 10.22587/rjabs.2023.15.1.2.
4. Al-Jebory, H.H., and S.A.H. Naji. 2021. Effect of Pelleted Fermented Feed in Production Performance of Laying Hens. *Fourth International Conference for Agricultural and Sustainability Sciences IOP Conf. Series: Earth and Environmental Science* 910 (2021) 012007 IOP Publishing doi:10.1088/1755-1315/910/1/012007.
5. AL-JEBORY, H.H., H.F.A.-H. AL-JEWAHERY, MKI. AL-SAEEDI, I.M.A Abd ALZAHRA, I.H. KADEM, N.A.-L. ALI. 2024. The Antibacterial effects and chicks quality response of copper nanoparticles of Japanese quail in hatching. *Mor. J. Agri. Sci.* 5 (1): 40-44.
6. Aljebory, H.H.D., and S.A.H. Naji. 2021. Effect of Pelleted Fermented Feed-in Egg Quality of Laying Hens. *Diyala Agricultural Sciences Journal* 13 (1): 41-57. <https://dx.doi.org/10.52951/dasj.21130105>.
7. Al-Saeedi, M.k.i., H.H. Al-Jebory and ,M.H. Abood. 2022. Progress Phenotypic Traits of Hatched Chicks and Growth Indicators of Broiler Chicks Fed Embryonically with Zinc Methionine. *Archives of Razi Institute.* 77(6): 2139-2145.
8. AL-SAEEDI, MKI, HH AL-JEBORY and M. AJAFAR. 2023. Effect of in Ova Injection with Nano-copper in Productive Performance of Japanese Quail Exposed to Pathological and Environmental Challenges. *Annals of Agri-Bio Research* 28 (2): 361-366.
9. Bell, D. 1996. Effects of temperature and storage time on egg weight loss. *Poultry International* 35(14):56-64.
10. Carlander, D., M. Wilhelmson, and A. Larsson .2003. Immunoglobulin Y Levels in Egg Yolk From Three Chicken Genotypes. *Food and Agricultural Immunology.* 15:1, 35-40, DOI: 10.1080/0954010031000138087.
11. Jones, D. R., Tharrington, J. B., Curtis, P. A., Anderson, K. E., Keener, K. M. and Jones, F. T. 2002. Effects of cryogenic cooling of shell eggs on egg quality. *Poultry Science* 81(5):727-733.
12. Kadhim, A.H., H.H. Al-Jebory, M.A. Ali and F.R. Al-Khafaji. 2021. Effect of Early Feeding (in Ovo) With NanoSelenium and Vitamin E on Body Weight and Glycogen Level in Broiler Chickens Exposed to Fasting Condition. *Fourth International Conference for Agricultural and Sustainability Sciences IOP Conf. Series: Earth and Environmental Science* 910. 012009 IOP Publishing doi:10.1088/1755-1315/910/1/012009.
13. Morais, C. F. A., Campos, E. J. and Silva, T. J. P. 1997. Qualidade interna de ovos comercializados em diferentes supermercados na cidade de Uberlândia. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia* 49:365-373.
14. Moula, N., Antoine-Moussiaux, N., Famir, F. and Leroy, P. 2009. Comparison of egg composition and conservation ability in two Belgian local breeds and one commercial strain. *International Journal of Poultry Science* 8(8):768-774.
15. Samli, H. E., Agna, A. and Senkoylu, N. 2005. Effects of storage time and temperature on egg quality in old laying hens. *Journal of Applied Poultry Research* 14:548-533.
16. Samli, H. E., Agna, A. and Senkoylu, N. 2005. Effects of storage time and temperature on egg quality in old laying hens. *Journal of Applied Poultry Research* 14:548-533.
17. Scott, T. A. and Silversides, F. G. 2000. The effect of storage and strain of hen on egg quality. *Poultry Science* 79(12):1725-1729.
18. Silversides, F. G. and Budgell, K. 2004. The relationships among measures of egg albumen height, pH and whipping volume. *Poultry Science* 83(10):1619-1623.
19. Walsh, T. J., Rizk, R. E. and Brake, J. 1995. Effects of temperature and carbon dioxide on albumen characteristics, weight loss, and early embryonic mortality of long stored hatching eggs. *Poultry Science* 74(9):1403-1410.
20. Zaki A.N., and HHD. Al-jebory. 2021. effect of early feeding with zinc-methionine on improving growth performance and some biochemical characteristics of broilers. *1st INTERNATIONAL VIRTUAL CONFERENCE OF ENVIRONMENTAL SCIENCES IOP Conf. Series: Earth and Environmental Science* 722 (2021) 012035 IOP Publishing doi:10.1088/1755-1315/722/1/012035.
21. Zeidler, G. 2002. Shell quality and preservation. In Bell, D. D. and Weaver, W. D. Jr. (eds). *Commercial Chicken Meat and Egg Production*. 5th rev. edn. Kluwer Academic Publishers, Norwell, pp. 1199-1217.