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Metabolite Profile of Moringa Leaves (*Moringa oleifera* Lam.) from Several Regions in South Sumatra, Indonesia

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ABSTRACT: Moringa (Moringa oleifera Lam.) is a medicinal plant which is recognized as having	Published Online:
many health benefits and has been widely used as a herbal medicine to treat certain diseases. To	14 June 2023
make an ingredient into herbal medicine, standardization of testing is required, because the chemical	
composition of a plant is not always the same. The growth of Moringa in South Sumatra is spread	
with varying altitudes, ranging from low, medium and high altitudes. Not much is known about the	
profile of metabolites of Moringa leaves based on the altitude where they grow. The altitude factor	
is one of the abiotic factors that can affect plant composition. The purpose of this study is to	
determine the metabolite profile and pharmacological activity of Moringa leaf metabolites based on	
different growing heights using a non-target metabolomics analysis approach using the GC-MS	
instrument. Sampling locations were determined using a stratified purposive sampling method based	
on the height where they grow. It is known that Moringa leaves from Bangun Rejo Village (795	
masl) and Masam Bulau Village (570 masl) showed 29 types of metabolites, while Tebing Gerinting	
Village, Ogan Ilir (35 masl) showed 27 types of metabolites. Each Moringa leaf growing at an	
altitude of 35 masl showed dominant metabolites Palmitic acid methyl ester and Linolenic acid	
methyl ester, a height of 570 masl with Linolenic acid methyl ester and Oleic acid methyl ester	
compounds and a height of 795 masl with Palmitic acid methyl ester compounds and Oleic acid	
methyl ester, which has bio-activity as an antioxidant, antiandrogenic, antiproliferative, antieczemic,	
antihistamine, antibacterial, antifungal, hypocholesteromiic and antitumor.	
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KEYWORDS: Altitude, GC-MS, Metabolite profile, <i>Moringa oleifera</i> Lam., Bio-activity.	Juswardi

INTRODUCTION

Herbal medicine has been widely accepted by the world community in dealing with various diseases. Knowledge, experience and skills regarding medicinal plants have been passed down from one generation to the next for generations. The World Health Organization (WHO) recommends herbal medicines for the maintenance of public health, because herbal medicines have relatively fewer side effects than modern medicines. The use of these herbal medicines must still pay attention to the accuracy of the dosage, the time of use, the method of use, the correctness of the drug, and the accuracy in choosing drugs for certain diseases (Sumayyah and Salsabilla, 2017).

Indonesia has great potential in the supply of herbal medicinal plants. *Moringa oleifera* Lam. Or Moringa with the local name Daun Kelor in Indonesia is a medicinal plant that is recognized as having many health benefits. The use of Moringa leaves can overcome several diseases, such as hyperglycemia, inflammation, bacterial or viral infections, and cancer (Tiloke *et al.*, 2018), as well as inhibit growth disorders caused by exposure to alcohol (Wardani and Suryono, 2019).

The results of the phytochemical tests of Moringa leaf extract showed compounds that act as drugs in treating certain diseases. The phytochemical content of Moringa leaf extract includes alkaloids, flavonoids, saponins, phenols, steroids/tripenoids, and tannins which have a role in maintaining health (Putra *et al.*, 2016). Oka *et al.* (2016) explained that the active compound hexane fraction of Moringa leaf powder extract with GC-MS contains 8 compounds that provide antioxidant effects and belong to the phenol group.

One of the analyzes that can be used to determine the composition of metabolites is by metabolomics analysis. Metabolomics is the total analysis of metabolite compounds in a sample and cells or tissues of an organism (Qi and Zhang, 2014). Metabolomics analysis is a powerful initial approach to determine the metabolite profile of a substance. One approach to

metabolomics analysis is untargeted metabolomics analysis which is widely used as a first step for more in-depth and detailed research. The aim of the analysis is to collect as much information as possible about the metabolites contained in biological samples (De Vos *et al.* 2007). Roessner and Beckles (2009), stated, metabolomics is the latest 'omics' technology with the aim of separating and measuring metabolite compounds resulting from plant biological activity.

Gas Chromatography-Mass Spectrometry (GC-MS) is one of the instruments used to identify metabolites in a material. The working principle of GC-MS is separation with high chromatography and being able to identify the metabolites of a material, such as essential oils, fatty acids, hydrocarbons, lipids, and others (Kaushik *et al.*, 2002). According to Sprakman *et al.* (2011) the advantage of GC-MS is that it has a high level of sensitivity, so it can be used to analyze compounds with small concentrations.

Moringa can grow in lowland areas to highlands, in range 0-1000 masl (Krisnandi, 2014). The composition of Moringa leaf metabolites is thought to be different at different heights where it grows. The growth of Moringa in South Sumatra is spread with varying altitudes, ranging from low, medium and high lands. Therefore, research is needed regarding the profile of Moringa leaf metabolites originating from that area in South Sumatra, to be used as a source of data and information regarding the type and abundance of metabolite compounds, as well as the dominant compounds in Moringa leaf samples from different heights.

MATERIAL AND METHODS

Sampling of Moringa leaves was carried out in several places in South Sumatra, in Tebing Gerinting Village, Ogan Ilir (35 masl), Masam Bulau Village, Lahat (570 masl), and Bangun Rejo Village, Pagar Alam (795 masl) South Sumatra. Moringa leaves methanol extract were analyzed for profile of metabolite

Research procedure

Sampling

Sampling locations were determined using a stratified purposive sampling method based on altitude, at lowland, medium, and high.

Preparation and Making of Simplicia

The obtained Moringa leaves were separated from each stalk. Furthermore, washed with running water, then drained and in the oven with a temperature of 50° C for 3x24 hours. After drying, it was weighed again to determine the dry weight. Then it was mashed using a blender until it became simplicia powder and sifted through a 12 mesh sieve. The refined simplicia powder is put into a jar that has been labeled according to the sampling location.

Extraction

The simplicia powder used was 100 g and 500 ml methanol solvent or a ratio of 1: was macerated for 1x24 hours with methanol solvent which was placed in a dark room at room temperature. After maceration is complete, it is filtered using filter paper, then concentrated using a rotary evaporator at 50°C until a thick extract is obtained.

Analysis of Moringa Leaves Using GC-MS

Sample preparation was carried out using methanol solvent. Moringa leaf methanol extract which had been added with 10 ml of methanol was injected as much as 1μ l into the GC-MS according to the instrument method based on the work protocol of GC-MS TraceTM 1310 ISQ

Data Analysis

The results of the identification of metabolites were analyzed by descriptive quantitative analysis in the form of data tabulation. GC-MS results data is in the form of a chromatogram containing a graph complete with a list of detected chemical components, chemical structure, retention time, and area. The profile of metabolites detected in each sample was calculated by calculating the total type and abundance of metabolites. The total abundance of types of metabolites was determined by calculating all the abundances of types of metabolites in each sample of Moringa leaves. Furthermore, the dominant metabolite compound was determined based on the largest percentage of area, then a biosynthetic pathway was traced to the dominant compound using the PubChem, KEGG, ChEBI, PlantCyc, and Spectrabase websites.

RESULTS AND DISCUSSION

Research on Moringa leaf metabolite profiles from several regions in South Sumatra based on differences in altitude, is presented in Table 1 as follows.

Metabolite Compounds	Molecular Formulas	Bio-activities	References	Pres	of	
	Formulas			TG	MB	BR
2-Myristynoyl pantetheine	C25H44N2O5S	-	-	+	+	+
N-[5-(1-Cyano-2-furan-2-yl- vinyl)-[1,3,4]thiadiazol-2-yl]- benzamide	C16H10N4O2S	Antitumor	Holmes & Twentyman (1995)	-	-	+
Deoxyspergualin	C17H37N7O3	Antioxidant and anti-inflammatory	Eryanti et al. (2011)	+	-	-
Propanoic acid, 3-ethoxy-, ethyl ester	C7H14O3	-	-	-	+	+
Cyclopentanone, 2-methyl	C6H10O	-	-	+	-	-
2-(2 Butynyl)cyclohexanone	C10H14O	-	-	+	-	-
4-(2,4,4-Trimethyl-cyclohexa- 1,5-dienyl)-but-3-e n-2-one	C13H18O	-	-	-	+	-
Toluene, m-ethyl-	C9H12	-	-	+	-	-
Benzene, p-dichloro-	C6H14Cl2	-	-	+	-	-
Tertbutyloxyformamide, N- methyl-N-[4-(1-pyrrolidinyl)-2- butynyl]-	C14H24N2O2	-	-	-	-	+
(4-Carbomethoxy)benzyl p- toluate	C17H16O4	Antimicrobial, anesthetic, antioxidant, antiseptic, anticancer.	Mary dan Giri (2008)	-	-	+
Benzoic acid, 4-methyl, [4(methoxycarbonyl)phenyl] methyl ester	C17H16O4	-	-	-	+	-
Acetonitrile, (p-hydroxyphenyl)	C8H7NO	Drugs for disorder of the urinary system, antiasthmatics	sSubramanian <i>et al</i> y(2020)		-	+
17-Pentatriacontene	C35H70	-		-	+	-
Methyl 2-ethylhexyl phthalate	C17H24O4	-	-	-	+	-
1-Methoxy-2-propyl acetate	C6H12O3	-	-	-	-	+
Ethanol, 2-(9-octadecenyloxy)	C20H40O2	-	-	-	-	+
Octaethylene glycol monododecyl ether	C28H58O9	-	-	+	-	-
Metabolite Compounds	Molecular Formulas	Bio-activities	References	Prese	ence abolites	(
	- Stillard			TG	MR	BR

9,10-Secocholesta-5,7,10(19)- triene-3,24,25-triol,	C27H44O3	Vitamins, drugs for autoimmune disorders and bon diseases, antidermatitis, anti inflammatory	orBrintha <i>et al.</i> (2017) ne i-	+	+	+
Methyl 9-cis,11-trans- octadecadienoate	C19H38O2	-	-	+	+	-
9,12,15-Octadecatrienoic acid	C27H52O4Si2	-	-	-	+	-
Methyl N-(N- benzyloxycarbonyl-beta-l- aspartyl)-beta-d- glucosaminide	C19H26N2O10	Alzheimer's disease drug	Rajmoahamed <i>et al</i> (2017)	<i>!.</i> +	+	+
Linolenin, 1-mono	C21H36O4	-	-	+	-	-
Linolenic acid, methyl ester	C19H34O2	Hypocholesterolemi c, antiandrogenic antiacne, antihistamine, antieczemic, antiproliferative, antifungal	i Nishanthini et al c,(2014)	<i>!</i> .+	+	+
Oleic acid, 3-	C39H76O3	-	-	+	+	+
(octaaecyloxy)propyl ester Oleic acid methyl ester	C19H36O2	Antibacterial	Teh et al (2017)) _	_	+
Olele delli, methyl ester	017113002	antitumor	Pinto <i>et al.</i> (2017)	',		1
Oleic acid, eicosyl ester	C38H74O2	Antibakterial	Awa et al. (2012)	+	+	+
Methyl tetradecanoate	C15H30O2	Antibakterial	Chandrasekara <i>et al</i> (2011)	!.+	+	+
i-Propyl 12-methyl-tridecoate	C17H34O2	-	-	+	-	-
Palmitic acid, methyl ester	C17H34O2	Antioxidant, antiandrogenic	Kumar <i>et al.</i> (2010)	+	+	+
Palmitic acid, 2- (hexadecyloxy)ethyl ester	C34H68O3	C C		-	+	-
Palmitic acid, ethylene ester	C34H66O4	-	-	-	+	-
Methyl 11-hexadecenoate	C17H32O2	-	-	-	-	+
Methyl 10-methylundecanoate	C13H26O2	-	-	-	-	+
Methyl stearate	C19H38O2	Antioxidant, antidiarrheal, antiinflammatory, antimicrobial, anticancer	Abdel-Hady <i>et al</i> (2018)	!	-	+
Methyl isostearate	C19H38O2	-	-	+	+	-
<i>i-Propyl</i> 14-methyl- pentadecanoate	C19H38O2	-	-	+	-	-
Gentamicin a	C18H36N4O10	Antibakterial	Scholar (2007)	+	-	-

Metabolite Compounds	Molecular	Bio-activities	References	Presence		of	
	Formulas			TG	MP	DD	
D-Mannosamine	C6H13NO5	Antitumor	Onoda <i>et al.</i> (1985)	-	+	- DK	
Phenol, 2,4-bis(1,1- dimethylethyl)	C14H22O	Antimicrobial, antioxidant	Teresa <i>et al.</i> (2014)	+	-	-	
2-Methoxy-4-vinylphenol	C9H10O2	-	-	-	+	+	
Ethyl iso-allocholate	C26H44O5	-	-	+	+	+	
7,8-Epoxylanostan-11-ol, 3- acetoxy-	C32H54O4	Antimicrobial, anti inflammatory	-Zekeya <i>et al.</i> (2014)	+	+	+	
5á)Pregnane3,20á-diol	C28H43NO6	-	-	-	+	-	
Dasycarpidan-1-methanol, acetate (ester)	C20H26N2O2	Anti-inflammatory	Mohammad <i>et al</i> (2016)	.+	+	-	
Pyridine, 2-(1-methyl-2- pyrrolidinyl)	C10H14N2	-	-	-	+	-	
Dihydromorphine, di(trimehylsilyl) ether	C23H37NO3Si2	-	-	-	+	-	
Imidazolo	C19H15N3O	-	-	-	-	+	
4-(4-Chlorophenyl)-N-[(2E)- 1-methylpiperidinylidene]-5- isothiazolamine	C15H16CIN3S	-	-	-	-	+	
Phorbol 12,13-dihexanoate	C32H48O8	-	-	+	+	-	
Lycopene	C42H64O2	Antioxidant	Miller <i>et al.</i> (1996)	-	-	+	
Octadecane	C26H54	-	-	-	+	-	
Phytol	C20H40O	Antioxidant, antidiarrheal, antimicrobial, anticancer, antiinflammatory, antihyperalgesic, Antiarthritic	Carvalho <i>et al.</i> (2020), Abdel-Hady <i>et al.</i> (2018)	-	-	+	

Description: TG = Tebing Gerinting Village (45 masl), MB = Masam Bulau Village (570 masl), BR= Bangun Rejo Village (795 masl), minus sign (-) = not detected

The same metabolites detected in the Moringa leaf samples included 2-Myristynoyl pantetheine, 9,10-Secocholesta-7,10(19)-triene3,24, 25 -triol, Oleic acid eicosyl ester, Oleic acid 3-(octadecyloxy)propyl ester, Linolenic acid methyl ester, Methyl tetradecanoate, Palmitic acid methyl ester, Methyl N-(N-benzyloxycarbonyl-beta-l-aspartyl)-beta-d-glucosaminide, Ethyl iso-allocholate. and 7,8-Epoxylanostan-11-ol 3-acetoxy.

The same metabolite compounds were detected in samples of Moringa leaves from Tebing Gerinting Village (35 masl), Masam Bulau Village (570 masl), and Bangun Rejo Village (795 masl) showing several health benefits, namely as an antioxidant, antiandrogenic, anti-acne, antifungal, antieczemic, antihistamine, antibacterial, antimicrobial, antiproliferative, anti-inflammatory, antidermatitis, hypocholesterolemic, vitamins, drugs for autoimmune disorders, bone disease, and Alzheimer's (Table 1).

Moringa leaves from Tebing Gerinting Village (35 masl), Masam Bulau Village (570 masl), and Bangun Rejo Village (795 masl) each have unique metabolite compounds or compounds that were only detected in Moringa leaves from Tebing Gerinting

Village (35 masl). N The compounds are Deoxyspergualin, Cyclopentanone, 2-methyl, 2-(2 Butynyl)cyclohexanone, Toluene methyl-, Benzene p-dichloro, Octaethylene glycol monododecyl ether, Linolenin, 1-mono, i-Propyl 12-methyl-tridecoate, i-Propyl 14-methyl-pentadecanoate, Gentamicin a, Phenol, 2,4-bis(1,1-dimethylethyl), and Phthalic acid methyl tetradecyl ester. According to Teresa et al. (2014), Phenol, 2,4-bis(1,1-dimethylethyl) is a compound that can be used as an antimicrobial correlated with its antioxidant activity, and Gentamicin a according to Scholar (2007) has a function as a pharmacological agent that can slow down protein synthesis in bacteria.

Unique metabolite compounds in Moringa leaves from Masam Bulau Village (570 masl), namely 4-(2,4,4-Trimethylcyclohexa-1,5-dienyl)-but-3-e n-2-one, Benzoic acid, 4 -methyl, [4(methoxycarbonyl)phenyl]methyl ester, 17-Pentatriacontene, Methyl 2-ethylhexyl phthalate, 9,12,15-Octadecatrienoic acid, Palmitic acid 2-(hexadecyloxy)ethyl ester, Palmitic acid ethylene ester, Mannosamine, Octadecane , 5á) Pregnane3,20á-diol, Pyridine, 2-(1-methyl-2-pyrrolidinyl), and Dihydromorphine, di(trimehylsilyl) ether. According to Onoda et al. (1985), D-Mannosamine is a metabolite that is known to be used as an antitumor which combined with unsaturated fatty acids can provide a cytotoxic effect on T-lymphocyte cells.

The only metabolite detected in Moringa leaves from Bangun Rejo Village (795 masl) was N-[5-(1-Cyano-2-furan-2-yl-vinyl)-[1,3,4]thiadiazol-2-yl] -benzamide, Tertbutyloxyformamide, N-methyl-N- [4-(1-pyrrolidinyl)-2-butynyl]-, (4-Carbomethoxy)benzyl p-toluate, Acetonitrile, (p-hydroxyphenyl), 1-Methoxy-2- propyl acetate, 1,1-Cyclopropanedicarboxylic acid, 2-[2-cyano-1,1 bis(methoxycarbonyl)propyl dimethyl ester, Ethanol 2-(9-, octadecenyloxy), Methyl 11-hexadecenoate, Methyl 10-methylundecanoate, Methyl stearate, Methyl arachisate, Imidazolo, 4-(4-Chlorophenyl) -N-[(2E)-1 methylpiperidinylidene]5isothiazolamine, Lycopene, and Phytol. The unique metabolites identified as having bio-activity are Methyl stearate, Methyl arachisate, Lycopene, and Phytol while other unique compounds have not been identified. Phytol is a diterpene class compound which is only found in Bangun Rejo Village (795 masl). According to Mach (2015), Phytol is a degradation of chlorophyll which becomes a tocopherol or vitamin E precursor in plants with the help of the enzyme phytol kinase which phosphorylates phytol into phytyl-phosphate. Ogunlesi et al. (2009) stated, Phytol provides good prevention and a promising treatment against rheumatoid arthritis and possibly other chronic inflammatory diseases.

CONCLUSION

Moringa leaves from Tebing Gerinting Village (35 masl), Masam Bulau Village (570 masl), and Bangun Rejo Village (795 masl) show metabolite compounds which are known to have bio-activities such as antioxidant, antiandrogenic, anti-acne, antifungal, antieczemic, antihistamine, antibacterial, antimicrobial, antiproliferative, anti-inflammatory, antidermatitis, hypocholesterolemic, vitamins, medications for autoimmune disorders, bone disease, and Alzheimer's. Each sample of Moringa leaves showed metabolites which were only detected at one of the sampling locations and some of these metabolites are known to have health benefits.

SUGGESTION

Further research is needed regarding the effect of edapic factors on the metabolite profile of Moringa leaves based on altitude using GCMS analysis.

REFERENCES

- 1. Abdel-Hady, H. 2018. GC-MS Analysis, Antioxidant and Cytotoxic Activities of *Mentha spicata*. *European Journal of Medicinal Plants*. 26(1): 1-12. https://doi.org/10.9734/EJMP/2018/45751
- Awa, E.P., Ibrahim, S. and Ameh, D.A.2012. GC/MS Analysis and Antimicrobial Activity of Diethyl Ether Fraction of Methanolic Extract from the stem bark of *Annona senegalensis* Pers. Int J Pharm Sci Res. 3(11); 4213-4218. : https://doi.org/10.13040/IJPSR.0975-8232.3(11).4213-18
- 3. Britha, S., Rajesh, S., Renuka, R., Santhanakrishnan, VP. And Gnanam, R. 2017. Phytochemical analysis and bioactivity prediction of compounds in methanolic extract of *Curculigo orchioides* Gaertn. *Journal of Pharmacognosy and Phytochemistry*. 6(4): 192-197. https://www.phytojournal.com/archives/2017/vol6issue4/PartC/6-3-122-574.pdf
- Carvalho, A.M.S., Heimfarth, L., Pereira, E.W., Oliveira, F.S., Menezes, I.R., Coutinho, H., Picot, L., Antoniolli, A., Quintans, J., and Quintans-Junior, L. Phytol, a Chlorophyll Component, Produces Antihyperalgesic, Antiinflammatory, and Antiarthritic Effects: Possible NFκB Pathway Involvement and Reduced Levels of the Proinflammatory Cytokines TNF-α and IL-6. *Natural Products*. 83(4): 1107-1117. https://doi.org/10.1021/acs.jnatprod.9b01116
- 5. Chandrasekaran, M., Senthilkuma, A. and Venkatesalu, V. 2011. Antibacterial and antifungal efficacy of fatty acid methyl esters from the leaves of *Sesuvium portulacastrum* L. *Eur Rev Med Pharmacol Sci.* 15(7):775-80. https://www.europeanreview.org/wp/wp-content/uploads/995.pdf
- 6. De Vos RC, Moco S, Lommen A, Keurentjes JJ, Bino RJ, Hall RD. 2007. Untargeted large-scale plant metabolomics using liquid chromatography coupled to mass spectrometry. *Nat Protoc*. 2(4):778-91. https://doi.org/10.1038/nprot.2007.95

- Eriyanti, Y., Nurulita, Y., Hendra, R., Yuharmen, Y., Syahri, J. and Zamro, A. 2011. Synthesizing Derivatives From Cyclopentanone Analogue Curcumin And Their Toxic, Antioxidant And Anti-Inflammatory Activities. *Makara Sains*.15(2): 117-123. https://doi.org/10.7454/mss.v15i2.1060
- 8. Holmes, J. and Twentyman, P. 1995. The Activity of Deoxyspergualin in Multidrug-Resistant Cells. *Cancer Chemother Pharmacol*. 36(6): 499-505. https://doi.org/10.1007/bf00685800
- Kaushik, J.C., Sanjay, A., Tripathi, N.N., and Arya S. 2002. Antifungal Properties of Some Plant Extracts Against Damping Off Fungi of Forest Nurseries. *Indian Journal of Forestry*. 25:359-361. https://eurekamag.com/research/003/647/003647093.php
- 10. Krisnadi, A.D. 2014. Kelor Super Nutrisi. Blora: Kelorina.com. https://kelorina.com/ebook-kelor-super-nutrisi/
- 11. Kumar, P.S. Mishra, D., Ghosh, G., and Panda, C. 2010. Medicinal Uses and Pharmalogical Properties of *Moringa oleifera*. International Journal Phytomedicine. 2 : 210-216. https://ijp.arjournals.org/index.php/ijp/article/view/39/39
- 12. Mach J. 2015. Phytol from Degradation of Chlorophyll Feeds Biosynthesis of Tocopherols. *Plant Cell*. 27(10):2676 https://doi.org/10.1105%2Ftpc.15.00860
- 13. Mary, A. and Giri, R. 2018. GC-MS Analyisis of Bioactive Compounds of Achyranthes Aspera. World Journal of *Pharmaceutical Research*. 7(1): hhttps://doi.org/1045-1056. 10.20959/wjpr20181-10540
- 14. Miller, N. J., Sampson, J., Candeias, L. P., Bramley, P. M., and Rice-Evans, C. A. 1996. Antioxidant activities of carotenes and xanthophylls. *FEBS Letters*. 384(3): 240–242. https://doi.org/10.1016/0014-5793(96)00323-7
- 15. Mohammad, Ghaidaa & Al-Jassani, Mohammad & Hameed, Imad. (2016). Anti-bacterial, Antifungal Activity and Chemical Analysis of *Punica grantanum* (Pomegranate peel) Using GC-MS and FTIR Spectroscopy. International *Journal* of *Pharmacognosy and Phytochemical Research*. 8: 480-494. https://www.researchgate.net/publication/298294390_Antibacterial_Antifungal_Activity_and_Chemical_Analysis_of_Punica_grantanum_Pomegranate_peel_Using_GC-MS_and_FTIR_Spectroscopy
- Mukunzi, D., Nsor-Atindana, J., Xiaoming, Z., Gahungu, A., Karangwa, E., and Mukamurezi, G. 2011. Comparison of Volatile Profile of *Moringa oleifera* Leaves from Rwanda and China Using HS-SPME. *Pakistan Journal of Nutrition*. 10(7): 602-608.
- 17. Nishanthini, A., Mohan, V.R., and Jeeva, S. 2014. Phytochemical, FT-IR and GC-MS Analysis of Stem and Leaf of *Tiliacora acuminata* (Lan) Hook F and Thomas (Menispermaceae). *International Journal of Pharmaceutical Sciences and Reaserch*. 5(9): 3977-3986.
- 18. Ogunlesi, M., Okiei, W., Ofor, E., and Osibote, E. 2009. Analysis of the essential oil from the dried leaves of *Euphorbia hirta* Linn (Euphorbiaceae), a potential medication for asthma. *African Journal of Biotechnology*. 8 (24): 7042-7050.
- Oka, A.A., Wiyana, K.A., Sugitha, I.M., and Miwada, I.N.S. 2016. Identifikasi Sifat Fungsional Daun Jati, Kelor dan Kayu Manis dan Potensinya sebagai Sumber Antioksidan pada *Edible Film*. Jurnal Sain Peternakan Indonesia. 11(1):1-8. https://doi.org/10.31186/jspi.id.11.1.1-8
- 20. Onoda T, Morikawa S, Harada T, Morikawa K. 1985. Antitumor activity of D-mannosamine in vitro: cytotoxic effect produced by mannosamine in combination with free fatty acids on human leukemia T-cell lines. *Jpn J Clin Oncol*. 15(3):545-52.
- 21. Pinto, M.E., Araújo, S.G., Morais, M.I., Sá, N.P., Lima, C.M., Rosa, C.A., Siqueira, EP., Johann, S., and Lima. 2017. Antifungal and antioxidant activity of fatty acid methyl esters from vegetable oils. *An Acad Bras Cienc*. 89(3):1671-1681.
- 22. Putra, I.W., Dharmayudha, A.A., and Sudimartini, L.M. 2016. Identifikasi Senyawa Kimia Ekstrak Etanol Daun Kelor (*Moringa oleifera* L) di Bali. *Indomesia Medicus Veterinus*. 5(5):464-473.
- Qi, X., and Zhang, D. 2014. Plant Metabolomics and Metabolic Biology. *Journal of Integrative Plant Biology*. 56(9): 814-815.
- 24. Rajmohamed, M.A., Natarajan, S., Palanisamy, P., Abdulkader, A.M., and Govindaraju. 2017. Antioxidant and cholinesterase inhibitory activities of ethyl acetate extract of *Terminalia chebula*: Cell-free *In vitro* and *In silico* studies. *Pharmacognosy Magazine*. 13(51): 437-445.
- 25. Roessner, U., and Beckles. D.M. 2009. Metabolite Measurements. J. Schwender (ed.). *Plant Metabolic Networks*. New York: Springer.
- 26. Scholar, E. 2007. Gentamicin. Omaha : University of Nebraska Medical Center.
- 27. Sparkman, D., Penton, Z.E., and Kitson, F.G. 2011. *Gas Chromatography and Mass Spectrometry, Second Edition*. USA: Elsevier Inc.
- 28. Subramanian, S., Dowlath, M.J., Karuppannan, S.K., Saravanan, and Arunachalam, K.D., 2020. Effect of Solvent on the Phytochemical Extraction and GC-MS Analysis of Gymnema sylvestre. *Pharmacogn J.* 12(4):749-761.
- 29. Sumayyah, S. dan Salsabilla, N.2017. Obat Tradisional : Antara Kha-siat dan Efek Sampingnya. *Majalah Farmasetika*. 2(5):1-4.

- Teh, C.H., Nazni, W.A., Nurulhusna, A.H., Norazah, A., and Lee, H.L. 2017. Determination of antibacterial activity and minimum inhibitory concentration of larval extract of fly via resazurin-based turbidometric assay. *BMC Microbiology*. 17(36):1-8.
- 31. Tiloke, C., Anand, K., Gengan, R.M., and Chuturgoon, A.A. 2018. *Moringo oleifera* and Their Phytonanoparticles: Potential antiproliferative Agents Againts Cancer. *Biomedicine and Pharmacotherapy*. 108:457-466.
- 32. Teresa, M. R.-C., Rosaura, V.-G., Elda, C.-M., and Ernesto, G.-P. 2014. The Avocado Defense Compound Phenol-2,4-bis (1,1-dimethylethyl) is Induced by Arachidonic Acid and Acts Via The Inhibition ff Hydrogen Peroxide Production by Pathogens. *Physiological and Molecular Plant Pathology*. 87: 32–41.
- 33. Wardani, I.K., and Suryono. 2019. Effect of *Moringa oleifera* (Lam) Leaf Extracts on Growth of Chicken Embryo Induced by Alkohol. *NurseLine Journal*. 4(1):61-67.
- 34. Zekeya, N., Kidkuli, A.W., and Chacha, M. 2014. Analysisi of Phytochemical Composition of *Bersama abyssinica* by Gas Chromatography-Mass Spectrometry. *Journal of Pharmacognosy and Phytochemistry*. 3(4):245-252.
- 35. Zhu, Y., Yin, Q., and Yang, Y. 2020. Comprehensive Investigation of *Moringa oleifera* from Different Regions by Simultaneous Determination of 11 Polyphenols Using UPLC-ESI-MS/MS. *Molecules*. 25(676) : 1-25.