
Survey of Plant Disease Prevalence on Okra (*Abelmoschus esculentus* (L.) Moench) at Rivers State University

Chimankpa W. Worlu¹, Chibuzor Njoku¹, Fortune N. C. Okogbule¹, Adedoyin A. Worlu²

¹Department of Plant Science and Biotechnology, Faculty of Science, Rivers State University, Port Harcourt, Rivers State, Nigeria.

²Department of Agricultural and Applied Economics, Faculty of Agriculture, Rivers State University, Port Harcourt, Rivers State, Nigeria.

ABSTRACT

Survey of disease incidence and severity on okra (*Abelmoschus esculentus* (L.) Moench) was carried out to determine the geographic distribution and status of the diseases throughout Rivers State University. Three fields of okra were randomly selected and varied numbers of okra plant were observed and visually identified some disease symptoms associated with okra plant. Visual assessment method was used to estimate the qualitative and quantitative lesion that manifested on the plant stem and leaf. Percentage Ratio Scale for Visual Estimate of plant disease incidence and severity was adopted. Where the number of leaves/stems that were not diseased/infected was (0%), less than 10% infected (1), between 11-20% infected (3), between 21-39 infected (5), about 50-70% infected (7) and 71-100% infected (9). Inter-rating reliability/reproducibility of plant diseases between the observed fields was statistically determined using One-way t-Test analysis at 5% probability level. From the estimate, rust and ring spot occurred more frequently and had the highest percentage contribution of 14.0% each. Followed by leaf spot (13.1%), chlorosis and necrosis (9.2%), rot (8.3%), wilting (7.0%), mildew and scorch (6.7%) each, and curling (3.8%) which had the least percentage contribution of disease incidence in the fields. Result showed that disease severity/index of okra plants at the nearest minimum in the examined fields at Rivers States University was (25.2%).

Published Online: January 05, 2026

*Cite the Article: Chimankpa W. Worlu, Chibuzor Njoku, Fortune N. C. Okogbule Adedoyin A. Worlu (2026). Survey of Plant Disease Prevalence on Okra (*Abelmoschus esculentus* (L.) Moench) at Rivers State University. International Journal of Life Science and Agriculture Research, 5(1), 1-8. <https://doi.org/10.55677/ijlsar/V05I01Y2026-01>*

License: This is an open access article under the CC BY 4.0 license:

<https://creativecommons.org/licenses/by/4.0/>

Corresponding Author:

Chimankpa W. Worlu

KEY WORDS: Survey, Okra, Disease severity, Disease incidence, Reproducibility, Frequency

INTRODUCTION

Okra (Moench.) is considered to be one of the world's oldest crops and is cultivated in almost all the Inter-tropical and Mediterranean regions for its young fruits. The vegetable is an important source of vitamins and essential mineral salts including calcium, which lacks in the diet of poor people of most of the developing countries of the world. But the yield obtained in several parts of the world is relatively lower, for which there are many constraints including prevalence of diseases caused by different pathogens (Ahmad *et al.*, 2012; Arain *et al.*, 2012).

Okra is one of the most important and valuable vegetables of the tropical and subtropical areas (Khan *et al.*, 2005). In Nigeria okra is cultivated on several thousands of hectares but the yield obtained is not quite high, for which there are many constraints including prevalence of diseases caused by different biotic (pathogens) and abiotic (temperature, pH, soil moisture content, electrical conductivity, nutrients, etc) factors which are visibly seen and recognised as symptom of diseases (Shahid *et al.*, 2007; Mukhtar *et al.*, 2013).

Okra is susceptible to several diseases, both in the field and in storage (Aung and Prot, 1990; Floret and Serpantie, 1993; Rahoo *et al.*, 2011; Qureshi *et al.*, 2012; Vagelas and Gowen, 2012). Some varieties are highly susceptible to root decaying/root rot organisms

Chimankpa W. Worlu et al, Survey of Plant Disease Prevalence on Okra (*Abelmoschus esculentus* (L.) Moench) at Rivers State University

while some are associated with both field and storage deterioration of the fruits which results in yield loss (Khan *et al.*, 2005; Shahid *et al.*, 2007). This may however affect the economy leading to scarcity of income and food.

Quantification of plant diseases especially the intensity of disease symptoms on individual units (Severity) is the basis for several research in plant pathology and related disciplines, which include evaluating treatment effect, monitoring 'available resource and purpose – where accuracy is operationally defined as the closeness of the visual estimate to the actual value (Nutter *et al.*, 1991; Madden *et al.*, 2007).

Visual estimation is however referred to as the eye sensing stimulus (a diseased specimen), followed by perception of the sensation by our brains, which is in turn followed by a cognitive process based on our training, knowledge, and expertise to classify parts of the specimen as diseased (Madden *et al.*, 2007). Such fundamental process is sufficient to definitely ascertain incidence of disease, but more complex process is needed if an explicit estimate of severity based on the proportion of area diseased is to be made (Bock *et al.*, 2010).

Literatures have shown that about 70% of the populace of the world relies on agriculture and so, there is need for identification of the plant diseases in order to prevent losses within the yield (Khirade and Patil 2015). It is difficult and burdensome to observe plant diseases manually and would require enormous labour, expertise within the plant diseases, and excessive time interval (Madiwalar and Wyawahare 2017). Although sensor technology has been available to measure disease severity using the visible spectrum or other spectral range imaging, it is visual sensing and perception that still dominates, especially in field research (Moghadam *et al.*, 2017). One of the important and tedious tasks in agricultural practices is detection of disease on crops. It requires huge time as well as skilled labour (Shrestha *et al.*, 2020).

Diseases affect crop at all growth stages especially seedling stage, causing lesions and premature drying of young leaves which affect the panicle causing all kinds of physical manifestations (Getachew *et al.*, 2014). The fungi appear to overwinter as mycelia in the infected living leaves or dead plant debris in the soil and this can be influenced by high temperature, high relative humidity and leaf wetness are critical environmental factors that influence disease development ((Uddin, 2000; Ruiz, 2003). Disease incidence and severity in okra plant is of considerable economic importance and cause annual losses in tropics to an extent of 22 per cent (Hussain *et al.*, 2011; Kayani *et al.*, 2012).

Assessment and quantification of the incidence and severity of okra plant disease is important. This study is therefore aimed at surveying and assessing the incidence and severity of diseases in okra plant in order to determine the geographic distribution and status of the diseases throughout Rivers State University so as to prioritize research.

MATERIALS AND METHODS

Study Area and Sample Collection

A survey of okra fields located in different randomly selected localities of the major vegetable growing parts of Rivers State University was conducted for the determination of disease incidence and severity. The selected areas are the fields behind the department of Geology and opposite Faculty of Science, Rivers State University, Port Harcourt, Nigeria, which lies within latitudes $4^{\circ} 43'0743'07''$ and $4^{\circ} 54'3254'32''N$ and longitudes $6^{\circ} 56'0456'04$ and $7^{\circ} 03'2003'20''E$. The mean annual rainfall of the area is 2000mm and mean temperature of $29^{\circ} C$ (Tubonimi and Udonna 2015). From each locality (Geology and opposite Faculty of Science), three fields of okra were randomly selected and varied numbers of okra plant were selected, observed and visually identified some disease symptoms associated with okra plant from each of the fields.

Determination of Plant Disease Symptoms in Okra plant at Rivers State University

Visual assessment method was used to describe and estimate the qualitative and quantitative damage/lesion that manifested on the plant stem and leaf.

Percentage Disease Incidence in Okra Plant at Rivers State University

The Ratio Scale for Visual Estimate of plant disease incidence and severity based on percentage or proportion was adopted. That is, plant disease incidence was determined by measuring the proportion of diseased plants per treatment or the proportion of diseased leaves per plant (treatment) as representations of incidence (Campbell and Madden, 1990; Madden and Hughes, 1995; Waller *et al.*, 2002).

$$\text{Disease incidence (DI) expressed as} = \frac{\text{Number of infected plant parts}}{\text{Total number of plant assessed}} \times 100$$

Determination of Disease Severity in Okra Plant at Rivers State University

The Ordinal Scales (qualitative and quantitative) which descriptively defined the ranges of percentage scale was used to determine disease severity on the examined okra plants (Cainge *et al.*, 2020). The area or volume of plant tissue that was visibly diseased,

Chimankpa W. Worlu et al, Survey of Plant Disease Prevalence on Okra (*Abelmoschus esculentus* (L.) Moench) at Rivers State University

usually relative to the total plant tissues were measured, recorded and was presented in percentage of the relevant host tissues or organ covered by symptom or lesion or damaged by the disease (Campbell and Madden, 1990; Binns, 1994; Cooke, 2006). The number and size of the lesions were measured which however expressed the extent of damage caused by the pathogen (Mousanejad et al., 2010).

Rating was however done based on severity scale;

0: No disease on leaf/stem, 1: Small symptom covering <10% leaf/stem area, 3: Brown symptom 11-20% leaf/stem area, 5: spot/marginal symptom 21-39% leaf areas, 7: Circular brown sunken symptom 50-70% leaf/stem area and 9: Circular to irregular symptom 71-100% leaf/stem area.

Disease severity (DS)/Infection index =

$$\frac{\text{Sum of all disease rating}}{\text{Total no. of rating} \times \text{maximum disease grade}} \times 100$$

Determination of Frequency of Occurrence and Percentage Distribution of Diseases in Okra Plant at Rivers State University

Disease symptoms such as chlorosis, necrosis, leaf spot, rust wilting damping off, etc on Okra leaves and stems in the respective fields were individually counted, and the percentage distribution of the diseases was calculated for okra fields at Rivers State University, Port Harcourt, Nigeria.

$$\% \text{ Contribution} = \frac{\text{Number of occurred symptom/disease incidence}}{\text{Total number of symptoms/disease incidence}} \times 100$$

Determination of Reliability/Reproducibility of Plant Disease Prevalence/Data Analysis at Rivers State University

Inter-rating reliability/reproducibility of plant diseases between the observed fields was carried out through statistical analysis using One-way t-Test analysis at 5% probability level which determined the extent to which okra plants were infected with the respective diseases at Rivers State University were infected.

RESULTS

The result of this research is presented in four different headings; investigation of the type of fungal disease incidence and percentage disease incidence in okra plant, determination of the frequency of occurrence and percentage contribution of each of the disease symptoms in okra plant, determination of the percentage disease severity/index on okra plant, and determination of reliability/reproducibility of plant disease prevalence on okra plant at Rivers State University, Port Harcourt, Nigeria.

Disease Incidence and Percentage disease Incidence in Okra Plant at Rivers State University

Table 4.1 shows some common fungal diseases/symptoms and percentage disease incidence in okra plant at the fields behind Department of Geology, Faculty of Science, Rivers State University. Ring spot had the highest percentage incidence (71.1) followed by leaf spot (63.2), necrosis (37.1), damping-off (37.0), rot (36.4), rust (34.4), chlorosis (29.2), scorch (27.0), wilting (25.3), mildew (17.6) and curling (11.8) being the least incidence.

Table 1. Diseases/Symptoms and Percentage Disease Incidence on Okra Plant at the Field behind Department of Geology, Rivers State University

S/N	Infections/Symptoms	Number of plants/parts Assessed	Number of infected plants/parts	Not infected plants/parts	% Incidence	Disease
1	Chlorosis	1205	352	853	29.2	
2	Necrosis	1184	439	745	37.1	
3	Leaf spot	781	494	287	63.2	
4	Rust	723	249	474	34.4	
5	Wilting	826	209	617	25.3	
6	Mildew	1210	213	997	17.6	
7	Curling	671	79	592	11.8	
8	Ring spot	1014	721	298	71.1	
9	Damping-off	622	304	518	37.0	
10	Scorch	972	214	578	27.0	
11	Rot	857	312	545	36.4	

Chimankpa W. Worlu et al, Survey of Plant Disease Prevalence on Okra (*Abelmoschus esculentus* (L.) Moench) at Rivers State University

Table 2 shows some common fungal diseases/symptoms and percentage disease incidence in okra plant at the fields Opposite Faculty of Science, Rivers State University. Leaf spot (41.8) had the highest percentage incidence, followed by rust (40.1), mildew (28.2), rot (25.3), scorch (25.1), wilting (23.0), damping-off (22.2), chlorosis (22.0), ring spot (19.4), necrosis (16.8) while curling (13.1) had the least incidence.

Table 2. Diseases/Symptoms and Percentage Disease Incidence on Okra Plant at the Fields opposite Faculty of Science, Rivers State University

S/N	Infections/Symptoms	Number of plants/parts Assessed	Number of infected plants/parts	Not infected plants/parts	% Incidence	Disease
1	Chlorosis	1112	245	867	22.0	
2	Necrosis	913	153	760	16.8	
3	Leaf spot	843	352	491	41.8	
4	Rust	1025	411	614	40.1	
5	Wilting	892	205	687	23.0	
6	Mildew	772	218	554	28.2	
7	Curling	1271	167	1104	13.1	
8	Ring spot	940	182	758	19.4	
9	Damping-off	957	212	745	22.2	
10	Scorch	872	219	653	25.1	
11	Rot	875	221	654	25.3	

Determination of Frequency of occurrence and Percentage contribution of each of the Disease Symptoms on Okra Plant at Rivers State University

Table 3 shows the frequency of disease occurrence and percentage contribution of disease symptoms in the fields (Department of Geology and Opposite Faculty of Science, Rivers State University). From the estimate, rust and ring spot occurred more frequently and had the highest percentage contribution of 14.0 each. Followed by leaf spot (13.1), chlorosis and necrosis (9.2), rot (8.3), wilting (7.0), mildew and scorch (6.7) each, and curling (3.8) which had the least percentage contribution of disease incidence in the fields.

Table 3. Frequency of Disease Occurrence and Percentage Contribution of Disease Symptoms on Okra Plant at Rivers State University

S/N	Infections/Symptoms	Total number of Plants/Parts Assessed	Total number of Disease/Symptom Occurrence	% Contribution of Symptom
1	Chlorosis	2317	597	9.2
2	Necrosis	2097	592	9.2
3	Leaf spot	1624	846	13.1
4	Rust	1566	905	14.0
5	Wilting	1851	454	7.0
6	Mildew	1982	431	6.7
7	Curling	1942	246	3.8
8	Ring spot	1954	903	14.0
9	Damping-off	1779	516	8.0
10	Scorch	1664	433	6.7
11	Rot	1732	533	8.3
Total		20508	6456	100

Determination of the Percentage Disease Severity/Index on Okra Plant at Rivers State University

Table 4 shows the disease severity scale/disease index, where the number of leaves/stems that were not diseased/infected was (14391), less than 10% infected area (1291), between 11-20% infected (2562) parts, between 21-39 infected (861) parts, about 50-70% infected (1702) parts and 71-100% infected (40) parts. And the disease severity/index of okra plants in the examined field at Rivers States University was (25.2%).

Table 4. Determination of Percentage Disease Severity/Index in Okra Plant at Rivers State University

Scale/Grade	Number of rating	Disease rating	%Disease Severity/Index
0	6456	0	
1	1291	1291	
3	1562	1562	
5	861	4305	
7	1702	11914	25.2
9	1040	9360	
Total	6456	28432	

Determination of Reliability/Reproducibility of Plant Disease Prevalence on Okra at Rivers State University

Table 4.5 shows the reproducibility of plant disease prevalence on okra at RSU. From the table, chlorosis, necrosis, leaf spot, rust, curling, damping-off and rot had significant effect on okra plant while wilting, mildew and scorch had highly significant impact on okra plants at RSU

Table 5. Reproducibility of Plant Disease Prevalence at Rivers State University

Symptoms	Rate of infection	t-value	Sig.
Chlorosis	298.50±75.66	5.579	0.11*
Necrosis	296.00±202.23	2.070	0.28*
Leaf spot	423.00±100.40	5.958	0.10*
Rust	330.00±114.55	4.074	0.15*
Wilting	207.00±2.82	103.500	0.01**
Mildew	215.50±3.53	86.200	0.01**
Curling	122.50±62.93	2.753	0.22*
Ring spot	451.50±381.13	1.675	0.34*
Damping-off	258.00±65.05	5.609	0.11*
Scorch	216.50±3.53	86.600	0.01**
Rot	266.50±64.34	5.857	0.12*

DISCUSSION

Results from the survey showed variations in the prevalence, incidence and severity of okra in different farms at Rivers state University. Similar results were also reported by Khan *et al.* (2005) and Shahid *et al.* (2007). The results of these workers confirmed the present findings regarding the prevalence of certain symptoms of plant diseases occurring on some species of vegetables. It is clear from the results of these researchers that okra crop was the most susceptible host of plant diseases in vegetable growing areas. These variations in infections and infestations are attributed to many environmental and edaphic factors, as differences in various climatic and edaphic factors of this zone (RSU) have been found to be suitable for disease prevalence. There are reports which confirmed that distribution, prevalence, incidence and severity of fungi diseases are affected by varying agro-climatic conditions of the areas, soil type, moisture, soil pH and particular cropping sequence (Hussain *et al.*, 2011; Kayani *et al.*, 2012).

Okra plants are also influenced by the biological, chemical and physical characteristics of the soil environment (Mukhtar *et al.*, 2013; 2013a; 2013b). In Rivers State University, the prevalence of okra diseases is quiet low. This is because of higher amount of organic matter in the soil of the farms and also due to the fact that okra was sown in the soils which were fallow for the last few years (Sathish *et al.*, 2013). This fact is supported by the findings of other researchers who found that fallowing increased the organic matter contents of the soils and thereby reduced the number of nematodes and other soil pathogens (Aung and Prot, 1990; Floret and Serpantie, 1993; Rahoo *et al.*, 2011; Qureshi *et al.*, 2012; Vagelas and Gowen, 2012).

The frequency of occurrence and percentage contribution of rust and ring spot were found to be a bit higher from the research (Table 4.3). This high incidence is due to intense vegetable cropping pattern and the availability of suitable host throughout the year in these fields which allowed rapid multiplication of leaf spot and ring spot (Torkpo *et al.*, 2008; Nilesh 2012). Earlier a number of researchers reported that abundance of leaf spot and ring spot is highly dependent upon the presence of the suitable host plants (Cuc and Prot, 1992).

However, the relatively low incidence and severity of okra diseases at Rivers State University were also due to the cultivation of non-susceptible varieties, optimal temperature and comparatively adequate annual rainfall which has direct impact on the soil. These conditions would however not favour the multiplication, development and infection of okra plant. There are reports which showed

Chimankpa W. Worlu et al, Survey of Plant Disease Prevalence on Okra (*Abelmoschus esculentus* (L.) Moench) at Rivers State University

that plant disease spread and multiplication are influenced by soil type (Trudgill *et al.*, 2000; Das and Das, 2000; Bhosle *et al.*, 2004; Ravichandra and Krishnappa, 2004; Rathour *et al.*, 2006).

In this study, rust and ring spot had the highest frequency of disease incidence and percentage contribution of okra plant diseases. Next to these was leaf spot (Table 4.3) but disease severity was relatively low (Table 4.4) across the fields at RSU. This may be due to other factors of disease such as weather, host resistance, inoculum load and so on. The conditions that favour disease development differ slightly with the conditions that bring about the disease occurrence (Amadi *et al.*, 2009). A disease may occur but if the conditions are not right, its development could be halted due to some of those factors earlier mentioned. Fungi are spread primarily by spores which are produced in abundance (Amadi, 2005). The spores can be carried and disseminated by wind current, water (splashing or rain), soil (dust), insects, birds and the remains of plants that once were infected Oyetunji *et al.*, 2012). The extent of dissemination of the disease inoculums will determine the development and severity of the disease.

CONCLUSION

It is concluded from the present studies that okra is attacked by several abiotic and biotic factors which is predominantly manifested in rust and ring spot especially at Rivers State University. However, the disease severity/index of okra plants in the examined fields at Rivers States University was (25.2%).

REFERENCES

1. Ahmad, Z., Saifullah, F., Khan H. and Idrees, M. (2012). Chemical and biological control of *Fusarium* root rot of okra. *Pakistan Journal of Botany*, 44(1): 453-457.
2. Amadi, J.E. (2005). Endophytic and Exophytic Fungi Isolated from the seeds of *Tetracarpidium conophorum* in Ilorin, Nigeria. *Nigerian Journal of Pure & Applied Science*. 20:1757-1761.
3. Amadi, J.E. (2009). Studies on the fungi associated with soft rot of carrot *Daucus carota* in Ilorin Metropolis. *International Journal Tropical Agricultural and Food Systems* 33: 252- 254.
4. Amadi, J.E.; Adebola, M.O. and Eze, C.S. (2009). Isolation and Identification of a Bacterial blotch Organism from Watermelon *Citrullus lanatus* Thunb. Matsum. & Nakai. *Africa. Journal of Agricultural Research* .411: 1291-1294.
5. Amadi, J.E.; Salami, S.O. and Eze, C.S. (2010). Antifungal Properties and Phytochemical Screening of Different Extracts of African Basil *Ocimum gratissimum* L.. *Agriculture and Biol. Jour. of North America* 12: 163-166.
6. Arain, A.R., Jiskani, M.M., Wagan, K.H., Khuhro, S.N. and Khaskheli, M.I. (2012). Incidence and chemical control of okra leaf spot disease. *Pakistan Journal of Botany*, 44(5): 1769-1774.
7. Aung, T. and Prot, J.C. (1990). Effects of crop rotations on *Pratylenchus zae* and yield of rice cultivar UPL Ri-5. *Review Nematology*, 13: 445-448.
8. Bhosle, B.B., Mukesh, S. Puri, S.N. and Suvasish, D. (2004). Prevalence of phytophagous nematodes in rhizosphere of okra (*Abelmoschus esculentus* (L.) Moench.) in Parbhani District, Maharashtra, India. *Indian Journal of Nematology*, 34(1): 56-59.
9. Binns, M. R. (1994). Sequential Sampling for Classifying Pest Status. Pages 137-174 in: *Handbook of Sampling Methods for Arthropods in Agriculture*. L. P. Pedigo and G. D. Buntin, eds. CRC Press, Boca Raton, FL.
10. Bock C.H, Poole G.H, Parker P.E, and Gottwald T.R (2010). Plant disease severity estimated visually, by digital photography and image analysis, and by hyperspectral imaging. *Critical Reviews in Plant Science* 29:59–107.
11. Campbell, C. L. & Madden, L. V. (1990). *Introduction to Plant Disease Epidemiology*. Wiley Interscience, New York.
12. Cooke, B. M. (2006). Disease Assessment and Yield Loss. In: *The Epidemiology of Plant Diseases*, pp.43-75, (Cooke, B.M., Jones, D.G. and Kaye, B., eds). Springer, the Netherlands. Culture médium. Pesqui. Agropecu. Bras, 43(4), 529-535 (in Portuguese).
13. Cuc, N.T.T. and Prot, J.C. (1992). Effect of changing the agricultural environment on ufra occurrence in the Mekong delta. *International Rice Research Newsletter*, 17: 25.
14. Das, J. and Das, A.K. (2000). Prevalence of root-knot nematodes on vegetable crops in Assam and Arunachal Pradesh. *Indian Journal of Nematology*, 30(2): 244-245.
15. Floret, C. and Serpantie, G. (1993). La jache're en Afrique de l'Ouest. ORSTOM, Colloques et se'minaires, Paris.
16. Getachew, G., Tesfaye, A. and Kassahun, T. (2014). Evaluation of disease incidence and severity and yield loss of finger millet varieties and mycelial growth inhibition of *Pyricularia grisea* isolates using biological antagonists and fungicides in vitro condition. *Journal of Applied Biosciences* 73:5883– 5901
17. Hussain, M.A., Mukhtar, T and Kayani, M.Z. (2011). Assessment of the damage caused by *Meloidogyne incognita* on okra. *Journal of Animal and Plant Science.*, 21: 857-861.

18. Kayani, M.Z., Mukhtar T. and Hussain M. A.. (2012). Evaluation of nematicidal effects of *Cannabis sativa* L. and *Zanthoxylum alatum* Roxb. against root-knot nematodes, *Meloidogyne incognita*. *Crop Protection*, 39: 52-56.
19. Khan, H.U., Mukhtar T. and Ahmad R. (2005). Geographical distribution of root-knot nematodes (*Meloidogyne* spp.) in the Punjab Province of Pakistan. *Pakistan Journal of Nematology*, 23(1): 133-140.
20. Kharade S. D. and Patil, A. B. (2015). "Plant Disease Detection Using Image Processing," International Conference on Computing Communication Control and Automation, 768-771
21. Madden L.V., Hughes G., and van den Bosch F. (2007). The Study of Plant Disease Epidemics. APS Press, St. Paul, MN.
22. Madden, L. V., and Hughes, G. (1995). Plant disease incidence: Distributions, heterogeneity, and temporal analysis. *Annual Review of Phytopathology*. 33:529-564.
23. Madiwalar, S. C. and Wyawahare, M. V. (2017) "Plant disease identification: A comparative study," 2017 International Conference on Data Management, Analytics and Innovation (ICDMAI), 13-18
24. Moghadam, P., Ward, D., Goan, E., Jayawardena, S., Sikka, P. and Hernandez, E. (2017). "Plant Disease Detection Using Hyperspectral Imaging," International Conference on Digital Image Computing: Techniques and Applications (DICTA), 1-8
25. Mousanejad, S., Alizadeh, A. & Safaie, N, (2010). Assessment of Yield Loss due to Rice Blast Disease in Iran. *Journal of Agricultural Science and Technology*, 12, 357-364.
26. Mukhtar, T. Kayani M.Z. and Hussain M.A. (2013). Response of selected cucumber cultivars to *Meloidogyne incognita*. *Crop Protection*, 44: 13-17.
27. Mukhtar, T., Kayani M.Z. and. Hussain M.A. (2013a.) Nematicidal activities of *Cannabis sativa* L. and *Zanthoxylum alatum* Roxb. against *Meloidogyne incognita*. *Ind. Crop Production*, 42: 447-453.
28. Mukhtar, T., Kayani M.Z., and Hussain, M.A. (2013a.) Nematicidal activities of *Cannabis sativa* L. and *Zanthoxylum alatum* Roxb. against *Meloidogyne incognita*. *Indian Crop Production*, 42: 447-453.
29. Mukhtar, T., Kayani, M.Z. and Hussain, M.A. (2013b). Response of selected cucumber cultivars to *Meloidogyne incognita*. *Crop Protection*, 44: 13-17.
30. Niles J. (2012). A review on *abelmoschus esculentus*; *pharmacacia*, 1: 1-8.
31. Nutter, Jr., F. W., Gleason, M. L., Jenco, J. H., and Christians, N. C. (1993). Assessing the accuracy, intra-rater repeatability and inter-rater reliability of disease assessment systems. *Phytopathology*, 83:806-812.
32. Oyetunji, E.O., Amadi, J.E., Peluola, C.O., Nwilene, F.E. and Akinwale, G. (2012). Comparative Effects of Tomato Juice Agar TJA, Potato Dextrose Agar PDA, Corn Meal Agar CMA, and Malt Extract Agar MEA on the Vegetative Growth of Fungi Associated with Soft Rot of Tomato *Lycopersicon esculentum* Mills Fruits. *Nigerian Journal of Botany* 252: 225-231.
33. Qureshi, S., Ruqqia, A.V. Sultana, J. A. and Ehteshamul-Haque S. (2012). Nematicidal potential of culture filtrates of soil fungi associated with rhizosphere and rhizoplane of cultivated and wild plants. *Pakistan Journal of Botany*, 44(3): 1041-1046.
34. Rahoo, A.M., Mukhtar, T. Gowen, S.R. and Pembroke, B. (2011). Virulence of Entomopathogenic Bacteria *Xenorhabdus bovienii* and *Photorhabdus luminescens* against *Galleria mellonella* Larvae. *Pakistan Journal Zoology*, 43: 543-548.
35. Rathour, K.S., Jola P. and Sudershan G. (2006). Community structure of plant parasitic nematodes in Champawat District of Uttarakhand, India. *Indian Journal of Nematology*, 36(1):89-93.
36. Ravichandra, N.G. and Krishnappa, K. (2004). Prevalence and distribution of phytoparasitic nematodes associated with major vegetables crops in Mandya District, Karnataka. *Indian Journal of Nematology*, 34(1): 113-116.
37. Ruiz, C. P. (2003). A new means of control for *Pyricularia oryzae*, *Rhizoctonia solani*, and other important rice-disease pathogens in Colombia. *Pflanzenschutz-Nachrichten Bayer* 56: 399-416.
38. Sathish K. D, Eswar T. D., Praveen K. A. Ashok K., Bramha S. D., and Rao, R N. (2013). International Research Journal of Pharmaceutical and Applied Sciences (IRJPAS). Available online at www.irjpas.com International Research Journal *Pharmaceutical and Applied Science*. 3(4):129-132
39. Shahid, M., Rehman A.U., Khan A.U. and Mahmood. A. (2007). Geographical distribution and infestation of plant parasitic nematodes on vegetables and fruits in the Punjab province of Pakistan *Journal of Nematology* 25(1): 59-67.
40. Shrestha, G., Deepak, M. Das, N. and Dey, N. (2020). "Plant Disease Detection Using CNN," 2020 IEEE Applied Signal Processing Conference (ASPCON), 109-113
41. Torkpo, S.K., Danquah, E.Y., Offei, S.K., and Blay E.T. (2008). Esterase, total protein and seed storage protein diversity in okra. *West Africa journal of applied ecology*. 9: 8-18.
42. Tripathi, K.K., Govila, O.P., Warrier, R. and Ahuja, V. (2011). Biology of *Abelmoschus esculentus* L. Okra. Series of Crop Specific Biology Documents 35 p., Department of Biotechnology, Ministry of Science & Technology & Ministry of Environment and Forestry, Govt. of India.

43. Trivino, C.G. and Voyoukallou, E. (2000). The importance of tropical root-knot nematodes (*Meloidogyne* spp.) and factors affecting the utility of *Pasteuria penetrans* as a biocontrol agent. *Nematology*, 2(8): 823-845.
44. Trudgill, D.L.G., Bala, V.C. Blok, A., Daudi, A., Davies, K.G., Gowen, S.R., Fargette, M., Madulu, J.D., Mateille, T., Mwageni, W., Netscher, C., Phillips, M.S., Sawadogo, A.
45. Uddin W, (2000). Gray leaf spot comes on strong. [Online] available: http://groundsmag.com/ar/grounds_maintenance_gray_leaf_spot/ (09 Oct. 2008).
46. Tubonimi, J. K. I. & Udonna, I. (2015). Hydrogeochemical Characteristics and Quality Assessment of Groundwater in University of Science and Technology Port Harcourt *International Journal of Environmental Monitoring and Analysis* 3(4), 221-232.
47. Vagelas, I. and Gowen, S.R. (2012). Control of *Fusarium oxysporum* and root-knot nematodes (*Meloidogyne* spp.) with *Pseudomonas oryzihabitans*. *Pakistan Journal Phytopathology*, 24:32-38.
48. Waller, J. M., Lenne, J. M. & Waller. S. J. (2002). Plant Pathologist's Pocketbook. 3rd edn. CABI Publishing, New York. 27.